

8. Loeblich, V. M., Baldwin, D. E., O'Connor, R. T. and Lawrence, R. V. (1955) *J. Am. Chem. Soc.* **77**, 6311.
9. Gough, L. J. and Mills, J. S. (1972) *Nature* **239**, 527.
10. Schubert, K. (1961) *Beihefte zum Geologischen Jahrbuch*, 45.
11. Beck, C. W., Meret, S., Kossove, D. and Kermani, K. (1965) *Archaeometry* **8**, 96.
12. Rottlander, R. C. A. (1970) *Archaeometry* **12**, 35.
13. Langenheim, J. H. (1969) *Science* **163**, 1157.
14. Dauben, W. C. and German, V. F. (1966) *Tetrahedron* **22**, 679.
15. Ternier, H., Ternier, G. (1960) *Atlas de Paléogéographie* p. 30. Masson, Paris.
16. Pavari, A. (1956) *Monti e Boschi* **11-12**, 555.

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FOUR EUDESMANOLIDES FROM *MONTANOA FRUTESCENS**

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Key Word Index—*Montanoa frutescens*; Asteraceae; Heliantheae; eudesmanolides; sesquiterpene lactones.

Abstract—Chemical analysis of *Montanoa frutescens* afforded the known sesquiterpene lactones montafusins A and B, as well as four new eudesmanolides which we have named montafusins C, D, E and F. The structures of the new compounds were established by spectroscopic methods.

INTRODUCTION

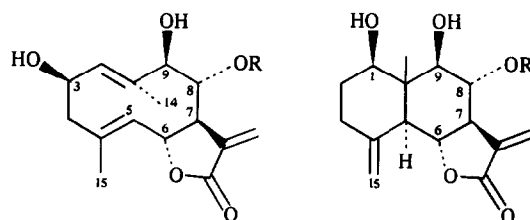
As part of our continuing study on the chemistry of plants of the genus *Montanoa*, we have examined the sesquiterpene lactones of the Mexican species *M. frutescens* [1], *M. tomentosa* [2, 3] and *M. grandiflora* [4].

Recently we re-investigated *M. frutescens* and isolated, besides the already described montafusins A (**1a**) and B (**1b**), four new eudesmanolides which we have named montafusins C, D, E and F.

RESULTS AND DISCUSSION

Montafusin C (**2a**), $C_{20}H_{26}O_6$, mp 140–144°, gave rise to two one-proton doublets at δ 6.14 ($J = 3.0$ Hz) and 5.48 ($J = 3.0$ Hz) and IR absorption at 1750 cm^{-1} typical of an α,β -unsaturated γ -lactone. Further IR absorptions at 1715 and 3400 cm^{-1} indicated the presence of an α,β -unsaturated ester and hydroxyl groups. The unsaturated ester was shown to be an angelate on the basis of the diagnostic ^1H NMR absorptions at δ 6.24, 2.05 and 1.95 due to the vinyl proton and methyl groups together with mass spectral peaks at m/z 83 and 55.

Detailed ^1H NMR spin-decoupling experiments allowed the major structural assignments of montafusin C (**2a**). Irradiation of the triplet of triplets at δ 2.91 (H-7)

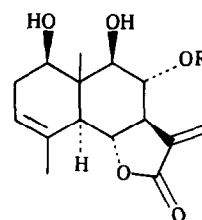


1a R = Ang

1b R = Sen

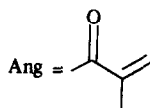
2a R = Ang

2b R = Sen

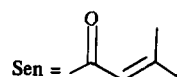


3a R = Ang

3b R = Sen



Ang =



Sen =

*Contribution No. 711 from Instituto de Química, UNAM, México.

collapsed the H-13 doublets at 6.14 and 5.48 to singlets and changed the doublet of doublets at 5.42 ($J = 11$, $J = 9.0$ Hz) to a doublet and the triplet at 4.16 ($J = 11.0$ Hz) to a doublet. The last two absorptions were assigned to the proton on the carbon carrying the ester group (H-8) and the proton at the lactonic carbon (H-6), respectively. In turn, irradiation at $\delta 4.16$ (H-6) changed the doublet at 2.26 ($J = 11.0$ Hz, H-5) to a broad singlet and the H-7 signal to a doublet of triplets. Saturation of the H-8 signal at $\delta 5.42$ collapsed the doublet at 3.89 ($J = 11.0$ Hz, H-9) to a broad singlet and the H-7 multiplet to a broad doublet. Irradiation at the frequency of H-9 and H-1 ($\delta 3.88$) collapsed the H-8 doublet of doublets at $\delta 5.42$ to a doublet and affected the one-proton multiplets centred at 1.84 (H-2a) and 1.55 (H-2b). Conversely, irradiation at the frequency of the H-2 multiplets affected the H-1 signal at $\delta 3.87$. The above spectral data established the structure **2a** for montafusurin C.

The stereochemistry of C-1, C-5, C-6, C-8 and C-9 followed from the couplings observed.

Montafusurins D (**2b**), E (**3a**) and F (**3b**) exhibited ^1H NMR (Table 1) and mass spectral signals which were nearly identical with those of montafusurin C (**2a**), except for absorptions that indicated differences in the ester group attached at C-8 and the double bond at C-4.

EXPERIMENTAL

M. frutescens (Maire) Hemsl. (2 kg) collected in Morelos, México, ca 60 km S. of México City in September 1981, was extracted as described before [1]. The CHCl_3 extract (85 g) was further fractionated on silica gel (800 g) using CHCl_3 and EtOAc. The EtOAc fraction (37 g) was chromatographed on silica gel (500 g) using petrol and petrol-EtOAc mixtures. Fractions eluted with petrol-EtOAc (17:3 and 7:3) were combined and rechromatographed on silica gel. Repeated TLC on fractions eluted with petrol-EtOAc (3:1 and 7:3) afforded the new eudesmanolides.

Montafusurin C (2a). $\text{C}_{20}\text{H}_{26}\text{O}_6$, mp 140–144° (Et₂O); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 209 (20 590); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400, 1750, 1715, 1640; EIMS (probe) m/z (rel. int.): 362 [M^+] ($\text{C}_{20}\text{H}_{26}\text{O}_6$) (1.98), 344 [$\text{M} - \text{H}_2\text{O}^+$] (0.5), 262 [$\text{M} - \text{AngOH}^+$] (1.0), 244 [$\text{M} - \text{H}_2\text{O} - \text{AngOH}^+$] (3.0), 226 [$\text{M} - 2\text{H}_2\text{O} - \text{AngOH}^+$] (7.9), 83 [$\text{C}_5\text{H}_7\text{O}^+$] (100.0), 55 [C_5H_7^+] (40.0).

Montafusurin D (2b). $\text{C}_{20}\text{H}_{26}\text{O}_6$, mp 219–221°; $[\alpha]_{\text{D}}^{20}$ (MeOH) +109.9°; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 213 (31 800); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3410, 1765, 1715, 1640; EIMS (probe) m/z (rel. int.): 362 [M^+] (0.02), 344 [$\text{M} - \text{H}_2\text{O}^+$] (0.06), 262 [$\text{M} - \text{C}_3\text{H}_8\text{O}_2^+$] (0.5), 244 [$\text{M} - \text{H}_2\text{O} - \text{C}_3\text{H}_8\text{O}_2^+$] (4.0), 226 [$\text{M} - 2\text{H}_2\text{O} - \text{C}_3\text{H}_8\text{O}_2^+$] (7.3), 83 [$\text{C}_5\text{H}_7\text{O}^+$] (100.0), 55 [C_5H_7^+] (14.5).

Montafusurin E (3a). $\text{C}_{20}\text{H}_{26}\text{O}_6$, mp 209–211° (Et₂O); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 206 (16 800); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3410, 1745, 1712, 1635; EIMS (probe) m/z (rel. int.): 362 [M^+] (0.5), 344 [M

Table 1. ^1H NMR data of montafusurins C (**2a**), D (**2b**), E (**3a**) and F (**3b**) (80 MHz, CDCl_3 , TMS as internal standard)

H	2a	2b	3a	3b
1	3.87 <i>dd</i>	3.85 <i>dd</i>	4.05 <i>dd</i>	4.02 <i>dd</i>
3	Obs.	Obs.	5.40 <i>s</i> (<i>br</i>)	5.40 <i>s</i> (<i>br</i>)
5	2.26 <i>d</i> (<i>br</i>)	2.24 <i>d</i> (<i>br</i>)	Obs.	Obs.
6	4.16 <i>t</i>	4.14 <i>t</i>	4.08 <i>t</i>	4.07 <i>t</i>
7	2.91 <i>tt</i>	2.85 <i>tt</i>	2.85 <i>tt</i>	2.83 <i>tt</i>
8	5.42 <i>dd</i>	5.33 <i>dd</i>	5.40 <i>dd</i>	5.40 <i>dd</i>
9	3.89 <i>d</i>	3.84 <i>d</i>	3.84 <i>d</i>	3.82 <i>d</i>
13a	6.14 <i>d</i>	6.13 <i>d</i>	6.12 <i>d</i>	6.12 <i>d</i>
13b	5.48 <i>d</i>	5.49 <i>d</i>	5.48 <i>d</i>	5.48 <i>d</i>
14	1.02 <i>s</i>	1.00 <i>s</i>	1.08 <i>s</i>	1.08 <i>s</i>
15a	5.09 <i>s</i> (<i>br</i>)	5.08 <i>s</i> (<i>br</i>)	1.86 <i>s</i> (<i>br</i>)	1.86 <i>s</i> (<i>br</i>)
15b	4.94 <i>s</i> (<i>br</i>)	4.92 <i>s</i> (<i>br</i>)		
OCOR	6.24 <i>qq</i>	5.78 <i>s</i> (<i>br</i>)	6.23 <i>qq</i>	5.78 <i>s</i> (<i>br</i>)
	2.05 <i>dq</i>	2.23 <i>d</i>	2.05 <i>dq</i>	2.23 <i>d</i>
	1.95 <i>quint.</i>	1.97 <i>d</i>	1.95 <i>quint.</i>	1.96 <i>d</i>

J (Hz): 1, 2 = 11; 1, 2' = 5; 5, 6 = 6, 7 = 7, 8 = 11; 8, 9 = 9; 7, 13a = 7, 13b = 3; compounds **3a**, **3b**: 1, 2 = 10; 1, 2' = 7.

$-\text{H}_2\text{O}^+$ (0.1), 262 [$\text{M} - \text{C}_3\text{H}_8\text{O}_2^+$] (1.6), 244 [$\text{M} - \text{H}_2\text{O} - \text{C}_3\text{H}_8\text{O}_2^+$] (2.3), 226 [$\text{M} - 2\text{H}_2\text{O} - \text{C}_3\text{H}_8\text{O}_2^+$] (1.5), 83 [$\text{C}_5\text{H}_7\text{O}^+$] (100.0), 55 [C_5H_7^+] (33.5).

Montafusurin F (3b). $\text{C}_{20}\text{H}_{26}\text{O}_6$, mp 200–202° (Et₂O); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 210 (25 600); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3420, 1765, 1710, 1635; EIMS (probe) m/z (rel. int.): 362 [M^+] (0.5), 344 [$\text{M} - \text{H}_2\text{O}^+$] (0.2), 262 [$\text{M} - \text{C}_3\text{H}_8\text{O}_2^+$] (2.0), 244 [$\text{M} - \text{H}_2\text{O} - \text{C}_3\text{H}_8\text{O}_2^+$] (3.2), 226 [$\text{M} - 2\text{H}_2\text{O} - \text{C}_3\text{H}_8\text{O}_2^+$] (3.5), 83 [$\text{C}_5\text{H}_7\text{O}^+$] (100.0), 55 [C_5H_7^+] (14.0).

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REFERENCES

- Quijano, L., Calderón, J. S., Gómez G., F. and Ríos, T. (1979) *Phytochemistry* **18**, 843.
- Quijano, L., Calderón, J. S., Gómez G., F. and Ríos, T. (1982) *Phytochemistry* **21**, 2041.
- Quijano, L., Gómez G., F., Calderón, J. S., López, P. J. and Ríos, T. (1984) *Phytochemistry* **23**, 125.
- Quijano, L., Calderón, J. S., Gómez G., F., López, P. J., Ríos, T. and Fronczek, F. R. (1984) *Phytochemistry* **23**, 1971.